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QUANTITATIVE ESTIMATION OF URINARY METABOLITES IN RUMINANTS BY HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY

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ABSTRACT

A reversed-phase high-performance liquid chromatographic method for a simultaneous determination of creatinine (C); Uric Acid (A); Hipoxanthine (HX); Xanthine (X); Hippuric Acid (HA); Benzoic Acid (BA) and Phenylacetic Acid (PAA) in urine of ruminants is described.

Separation was achieved on a Nova-PaK column by gradient elution. Quantitation and detection limits were determined. Detection is effected by UV absorption at 254 nm with a total analysis time of less than 30 min. An aliquot of diluted urine is injected directly into the liquid chromatographic column.

The proposed HPLC method was verified for linearity, accuracy, precision and applicability.

INTRODUCTION

A series of compounds presents in the urinary excretion of ruminants were proposed recently as index of their nutritive status (1) (2), to establish

relationship between energy and N intake, requirements and reserves disponibility in each phase of productive cycle.

In order to study these relationships, an analytical method is needed to determine the urinary metabolites. A series of papers (3-8) reported HPLC techniques for determination of some metabolites in biological fluids, have been published over the last few years.

This report described a rapid HPLC method for the simultaneously determination of seven metabolites presents in the sheep urine to stablish profiles and mathematical relationships which allow to define the nutritive status.

MATERIAL AND METHODS

Instruments

The analyses were performed on a Waters 600E equipped with a Waters 484, UV Detector. Separation was acomplished on a Nova-PaK C18 (3.9 x 150 mm) and the quantitation was based on integration of peak areas using a Waters 745B Integrator.

Chemicals

All reagents were Analytical Reagent Grade. Standards were purchased from Merck and used without further purification.

Methanol, HPLC grade, was purchased from Carlo Erba. Water was previously distilled and purified with a Milli-RO 15 Reagent Grade Water System (Millipore).

Urine samples

Urine samples were centrifuged and filtered through Millipore (0.45 μm) filter and diluted 10-fold (or more when the concentrations were high in samples) in distilled water. 20 μl of filtrate were injected directly to the column. The urine samples were stable for several weeks when stored at -20°C .

Standard Solutions

To prepare the standard curves, eight concentrations of standard solutions were made up in a range of concentrations from (10 to 100 $\mu\text{g}/\text{ml}$) for C; (2 to 100 $\mu\text{g}/\text{ml}$) for UA; (1 to 10 $\mu\text{g}/\text{ml}$) for HX; (2 to 10 $\mu\text{g}/\text{ml}$) for X; (10 to 100 $\mu\text{g}/\text{ml}$) for HA; (10 to 100 $\mu\text{g}/\text{ml}$) for BA and (50 to 200 $\mu\text{g}/\text{ml}$) for PAA. The stock solutions were stable when stored at 4°C until used.

The quantification were achieved by regression analysis of the peak areas of each analyte against concentration. Triplicate injections of each concentration were made.

Chromatographic conditions

HPLC separations were carried out using a 30 min linear gradient programmed from solvent A (0.025M aqueous sodium acetate, buffered to $\text{pH}=4.5$) to a mixture solvent A: methanol (75:25), v/v). The flow rate was maintained at 0.5 ml/min until the first 10 min and then increased to 1 ml/min until the end of analysis.

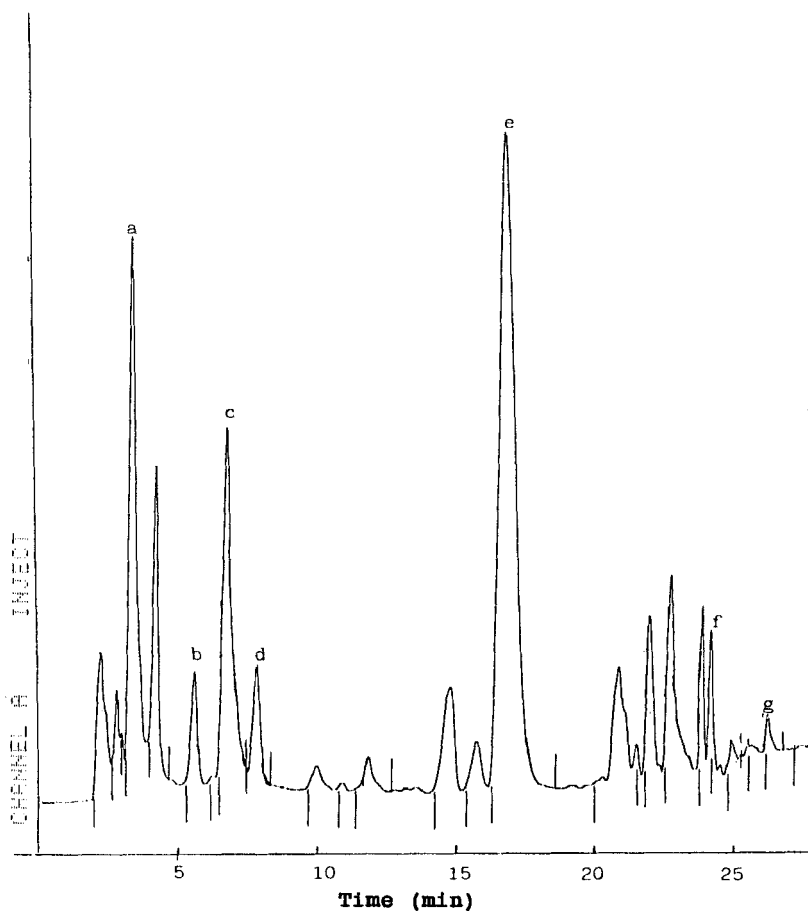


FIGURE 1.-Chromatogram of a ten-fold diluted sheep urine. Peaks: a = C; b = UA; c = HX; d = X; e = HA; f = BA and g = PAA.

Detector was set at 254 nm.

Sample volumes were 20 μ l, which were injected into the column through a Model U6K Injector (Waters Assoc.).

All analyses were carried out at room temperature.

The purity of the compounds peaks was tested by comparison of the peak areas obtained at wavelengths

of 245 and 254 nm from aqueous standards with those of various samples.

RESULTS AND DISCUSSION

Representative chromatogram obtained for urine samples is shown in Fig-1.

The retention times for C; UA; HX; X; HA; BA and PAA were approximately: 3.5; 5.4; 6.5; 7.3; 17.0; 23.8 and 26.0 min, respectively.

Calibration plots were linear over the concentrations range studied for the seven compounds. In all cases the correlation coefficients were found to be greater than 0.99. Fig-2 shows a typical calibration curve for HX.

The standard addition method was used to check for chemical interferences in the quantitation of different products. This method for HX in urine is shown in Fig-3. The slope found is similar to the calibration curve (Fig-2).

The limits of detection for ten-fold diluted urine were 1.0; 0.2; 0.1; 0.1; 0.7; 1.0; 7.0 $\mu\text{g/ml}$ for C; UA; HX; X; HA; BA and PAA with a 20 $\mu\text{l/ml}$ injection.

The absolute recoveries of the seven analytes from urine were assessed by comparing peak areas obtained for the standard stock solutions of the analytes and urine spiked with the respective analytes. In all cases, the value was estimated by subtracting the peak area obtained before spiking from the one found for spiked urine samples. Recoveries are shown in table I.

The interday precision, accuracy and reproducibility of the method over the entire concentrations range were determined with the analysis of spiked urine samples. As shown in table I, the

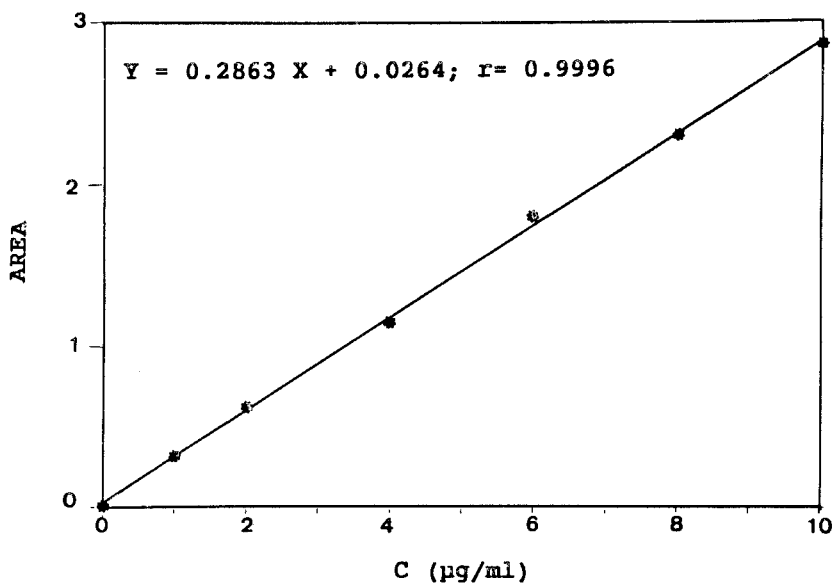


FIGURE 2.-Hipoxanthine calibration curve.

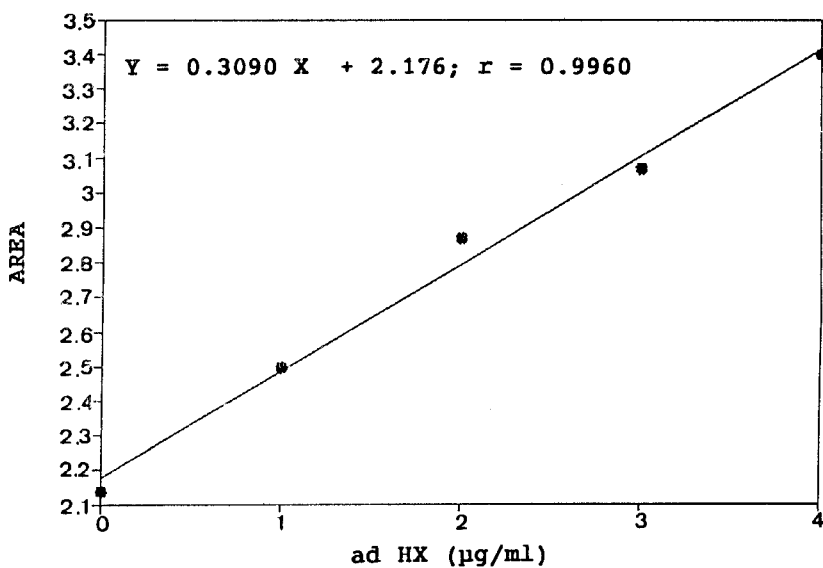


FIGURE 3.-Standard addition method for Hipoxanthine in sheep urine.

TABLE 1

Precision, Accuracy and Recoveries in the Determination of Metabolites in Sheep Urine

Comp. n=5	Conc. Added. ($\mu\text{g/ml}$)	Conc. found ($\mu\text{g/ml}$)	CV* (%)	RE** (%)	Recoveries Mean \pm SD (%)
Creati- nine	10	10.87 \pm 0.34	3.2	8.7	100.44 \pm 5.68
	20	19.73 \pm 0.43	2.2	1.4	
	30	29.37 \pm 0.89	3.0	2.1	
	40	38.60 \pm 1.28	3.3	3.5	
Uric Acid	2	2.03 \pm 0.04	2.0	1.5	101.24 \pm 2.03
	3	3.08 \pm 0.10	3.2	2.7	
	4	4.01 \pm 0.02	0.5	0.2	
	5	5.03 \pm 0.03	0.6	0.6	
Hipoxan- thine	1	0.99 \pm 0.02	1.6	1.0	100.18 \pm 1.71
	2	2.02 \pm 0.04	2.0	1.0	
	3	3.01 \pm 0.04	1.5	0.3	
	4	4.02 \pm 0.06	1.5	0.5	
Xanthine	1	0.98 \pm 0.03	2.8	2.0	99.95 \pm 1.87
	2	2.01 \pm 0.03	1.5	0.5	
	3	3.00 \pm 0.05	1.6	0.1	
	4	4.02 \pm 0.05	1.3	0.5	
Hippuric Acid	10	10.42 \pm 0.46	4.4	4.2	102.21 \pm 3.70
	20	20.85 \pm 0.83	3.9	4.2	
	30	30.23 \pm 0.44	1.4	0.8	
	40	39.82 \pm 0.48	1.2	0.4	
Benzoic Acid	20	20.85 \pm 0.86	4.2	4.2	101.22 \pm 3.10
	30	29.83 \pm 0.80	2.7	0.6	
	40	40.46 \pm 0.59	1.5	1.2	
	50	50.08 \pm 0.58	1.2	0.2	
Phenyl- acetic acid	150	147.99 \pm 2.73	1.8	1.3	99.50 \pm 1.50
	300	303.36 \pm 3.66	1.2	1.1	
	450	446.94 \pm 6.66	1.5	0.7	
	600	593.52 \pm 8.75	1.5	1.1	

* CV: Coefficient of variation; ** RE: Relative error

precision is expressed as the coefficient of variation and the accuracy as the relative error. For each analyte, interday precision was determined by assaying in 5 occasions, diluted urine samples spiked at 4 concentrations. In all instances, the coefficient of variation of peak area was found to be acceptable.

The accuracy was assessed by analysing known amounts of analytes. The observed concentrations were in good agreement with the actual concentrations.

In conclusion, we have developed a sensitive HPLC assay for the specific determination of Creatinine; Uric Acid; Hipoxanthine; Xanthine; Hippuric Acid; Benzoic Acid and Phenylacetic Acid in sheep urine, with high precision and accuracy. The method is also simple and rapid and suitable for routine analysis in clinical laboratories.

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REFERENCES

- 1.-ORSKOV, E.R., MacLEOD, N.A., FAHMY, S.T.M., ISTASSE, L and HOVELL, F D.D..-"Investigation of Nitrogen Balance in Dairy Cows and Steers Nourished by intragastric Infusion. Effects of Submaintenance Energy Input with or without Protein". Br. J. Nutr. 50, 99, (1983).
- 2.-HOVELL, F.D.D., ORSKOV, E. R., GRUBB, D. A. and MacLEOD, N. A. .-"Basal Urinary Nitrogen Excretion and Growth Response to Supplemental Protein by Lambs Close to Energy Equilibrium". Br. J. Nutr. 50, 173, (1983).

- 3.-VEENING, H.-"HPLC as Clinical Method for Monitoring Hemodialysis of Renal Patients". - Biological/Biomedical Applications of Liquid Chromatography II. eds, Marcel Dekker, New-York, 1979, p 93.
- 4.-SCHWEINSBERG, P. D. and LOO, T. L.-"Simultaneous Analysis of ATP, ADP, AMP and Other Purines in Human Erythrocytes by HPLC". J. Chromatogr. 181, 103, (1980).
- 5.-DEL RAZO, L.M. and JAUGE, P. .-"Quantitation of Creatinine in Urine and Plasma Sample by Reversed Phase HPLC". J. Liq. Chromatogr. 8(10), 1893, (1985).
- 6.-REGNAUD, L., SIROIS, G., COLIN, D. and CHAKRABARTY, S..-"Simultaneous Ion-Pairing Liquid Chromatographic Determination of the Major Metabolites of Styrene and Carbamazepine and of Unchanged Carbamazepine in Urine". J. Liquid Chromatogr. 10(11), 2369, (1987).
- 7.-XUE, G.P., FISHLOCK, R. C. and NOSWELL, A. M..-"Determination of Creatinine in Whole Blood, Plasma and Urine by High-Performance Liquid Chromatography". Anal. Biochem. 171, 135, (1988).
- 8.-KUBOTA, K., HORAI, Y, KUSHIDA, K. and ISHIZAKI, T..-"Determination of Benzoic Acid and Hippuric Acid in Human Plasma and Urine by HPLC". J. Chromatogr. 425, 67, (1988).